

**Supplementary Table S1.** Current information on the main effect of agricultural practices on mycotoxin content in maize.

Agricultural Practice	AFs	FBs	DON	ZEN	Notes	References
Sowing/plant density					FB1 and B2 occurrence was always significantly higher ( $P < 0.05$ ) with late (May) compared to early sowing date (March-April).	
		+	+		In DON, the effect of early sowing was only significant in 2007 for the early hybrid, while the effect of high density (80,000 pt/ha) vs low density (65,000 pt/ha) differed over the three years according to the hybrid maturity.	[1]
					Earlier planting consistently resulted in lower ear rot severity and FB1 contamination.	[2]
					High plant density (80000 pt ha <sup>-1</sup> ), nitrogen fertilization and late sowing date (May 3 to May 16) increased FBs contamination (+133%).	[3]
					Maize fields subjected to dry planting contained significantly more FBs ( $1,004 \pm 379$ $\mu\text{g}/\text{kg}$ ) than those sown by wet planting ( $230 \pm 88$ $\mu\text{g}/\text{kg}$ ) ( $P < 0.10$ ).	[4]
Hybrid					FBs concentration did not differ significantly depending on the hybrid season length. A significant increased DON concentration in the kernels when associated with the use of late maturity hybrids.	[1]
					Hybrid frequently had significant effects ( $P \leq 0.05$ ) on FB1 contamination.	[2]
					The effect of hybrid maturity and the interactions between the independent variables (agricultural practices and hybrid) and random factors (year and site) were never significant.	[3]
					The main role of fatty acids, with a higher FBs (FB1+FB2+FB3) contamination in hybrids showing a higher linoleic acid content and a higher masking action in hybrids with higher oleic to linoleic ratio.	[5]
Soil management					In the soil under no-tillage, contamination with fungal spores was 92.9% higher compared to the soil under conventional tillage. DON and ZEN content varied but it was not considerably influenced by the different tillage systems applied.	[6]
Irrigation					Drought level represented by weekly-ARID (Agricultural Reference Index for Drought) values before and after mid-silk is a significant predictor ( $p$ -value $< 0.10$ ) for AF contamination risk.	[7]
					Aridity indexes based on meteorological data confirmed the influence of drought conditions on AFB1 synthesis.	[8]
					AF levels averaged over a period of three years from $33.5 \pm 12.0$ $\mu\text{g}\cdot\text{kg}^{-1}$ for non-irrigated, $29.3 \pm 11.4$ $\mu\text{g}\cdot\text{kg}^{-1}$ for moderately irrigated and $24.2 \pm 8.5$ $\mu\text{g}\cdot\text{kg}^{-1}$ for well-irrigated plots.	[9]
					Irrigation significantly affected the level of FB contamination ( $P < 0.05$ )	[10]
					The type of irrigation had no distinct effect ( $P > 0.10$ ) on the FB levels, which were $508 \pm 219$ and $555 \pm 275$ $\mu\text{g}/\text{kg}$ in fields with flood and sprinkler irrigation systems, respectively.	[4]
					3.5 - to 5-times higher DON concentration in kernels at limited- than well-watered conditions: 380 compared with 75 $\mu\text{g}/\text{kg}$ .	[11]
Pesticides					The insecticide treatments (alpha-cypermethrin) against second-generation ECB larvae significantly reduce the FBs contamination but did not significantly reduce the DON contamination.	[1]
					Insecticide treatments (active ingredient: alpha-cypermethrin) reduced (-54%) the FBs occurrence compared to the untreated control.	[3]
					ECB control (deltamethrin) and tebuconazole applied at BBCH67. FB1 varied from 6767 $\mu\text{g}/\text{kg}$ in the unsprayed plot to 4429 (-35%) and 3013 (-56%) $\mu\text{g}/\text{kg}$ in respectively the plot treated with tebuconazole and with the addition of deltamethrin.	[12]
					The efficacy of ECB control was evaluated at 89.96% for FBs with insecticide (deltamethrin) and 89.97% with insecticide+fungicide (deltamethrin and tebuconazole). The efficacy was evaluated at 85.40% for ZEN with insecticide and 82.10% with insecticide+fungicide.	[13]

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Supplementary Table S1. (Continued).

Agricultural Practice	AFs	FBs	DON	ZEN	Notes	References
			+		Lambda-cyhalothrin or deltamethrin (20 g ha <sup>-1</sup> ). DON levels were highly significantly affected by insecticide treatment ( $F_{1,88}=35.925$ ; $P < 10^{-4}$ ). On average, DON levels were significantly lower (151.73 µg/kg) in treated maize than in the control (849.04 µg/kg).	[14]
		+			Insecticides: A mixture of chlorpyrifos and cypermethrin applied at 0.450 and 0.045 kg ha <sup>-1</sup> respectively. A significant effect ( $P < 0.001$ ) of insecticide application timing on fumonisin occurrence in maize kernels was observed. Efficacy of the best application timing to control fumonisin occurrence was 73% in 2006 and 84% in 2007.	[15]

The agricultural practice has an impact (+); the agricultural practice does not have an impact (-)

Abbreviations: aflatoxins (AFs), fumonisins (FBs), deoxynivalenol (DON), zearalenone (ZEN)

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**Supplementary Table S2.** Current information on reduction of mycotoxin-producing *Aspergillus* spp and *Fusarium* spp growth and mycotoxin production by plant-produced compounds.

Plant (compound)	Target fungal species	Type of assay	Reduction	References
<b>Essential oils (EOs)</b>				
<i>Eucalyptus grandis</i> , <i>E. staigeiriana</i> , <i>E. citriodora</i> , and the hybrid <i>E. grandis</i> x <i>E. urophylla</i>	<i>A. flavus</i> <i>A. parasiticus</i>	<i>In vitro</i>	<i>E. staigeiriana</i> showed the best potential on fungal growth control. The major active ingredients were limonene and geranial. The effect on mycotoxin production was not tested.	[1]
<i>Cymbopogon citratus</i>	<i>A. flavus</i> <i>A. parasiticus</i> <i>A. ochraceus</i> <i>A. niger</i> <i>A. fumigatus</i>	<i>In vitro</i>	The antifungal activity tests showed that the oil was active against all the five <i>Aspergillus</i> species, and the minimum inhibitory concentration (MIC) of the oil ranged from 15 to 118 mg/ml. The MIC* ranged from 15 to 118 mg/ml. The major active ingredients were geranial, neral, myrecene and geraniol. The effect on mycotoxin production was not tested.	[2]
Thyme ( <i>Thymus vulgaris</i> ), rosemary ( <i>Rosmarinus officinalis</i> ), and laurel ( <i>Laurus nobilis</i> )	<i>A. flavus</i> <i>A. parasiticus</i>	<i>In vitro/in vivo</i>	Thyme showed the highest inhibition on <i>A. parasiticus</i> growth (39 mm diameter of inhibition zone), followed by rosemary (15 mm) and laurel (10 mm).	[3]
Clove ( <i>Syzygium aromaticum</i> ) and vatica ( <i>Vatica diospyroides</i> )	<i>A. flavus</i>	<i>In vitro/in vivo</i>	Clove showed 84.7% inhibition on conidial germination of <i>A. flavus</i> at 100 $\mu\text{L L}^{-1}$ , and complete inhibition of disease infection on maize seeds at 10 $\mu\text{L L}^{-1}$ . Vatica completely inhibited growth, sporulation, conidial germination, and disease infection of <i>A. flavus</i> both <i>in vitro</i> and on maize seeds at 50 $\mu\text{L L}^{-1}$ . The main active ingredients were eugenol (62.4%) and benzyl acetate (48.8%) for clove and vatica oil respectively. The effect on mycotoxin production was not tested.	[4]
Cinnamon ( <i>Cinnamomum verum</i> ) essential oil (CEO)	<i>A. flavus</i>	<i>In vitro</i>	<i>A. flavus</i> growth rate diminished and lag time increased as the concentration of CEO increased. The major active ingredients was cinnamaldehyde.	[5]
Cinnamaldehyde, citral and eugenol	<i>A. flavus</i>	<i>In vitro</i>	<i>A. flavus</i> growth and AFB <sub>1</sub> production were completely inhibited by 0.80 mmol/L of cinnamaldehyde and 2.80 mmol/L of citral. At lower concentration, cinnamaldehyde, eugenol, and citral significantly reduced AFB <sub>1</sub> production with inhibition rate of 68.9%, 95.4%, and 41.8%, respectively, while no effect on fungal growth.	[6]
<i>Ocimum sanctum</i> essential oil (OSEO)	<i>F. graminearum</i>	<i>In vitro</i>	MIC and minimum fungicidal concentration of OSEO were 1250 and 1800 $\mu\text{g/mL}$ , respectively. ZEN concentration was insignificant at 1500 $\mu\text{g/mL}$ concentration. The main active ingredients was eugenol (34.7%).	[7]
Rocket seeds ( <i>Eruca sativa</i> ), rosemary ( <i>Rosmarinus officinalis</i> ) and tea tree ( <i>Melaleuca alternifolia</i> )	<i>F. graminearum</i> <i>F. avenaceum</i> <i>F. semitectum</i> <i>F. solani</i> <i>F. oxysporum</i>	<i>In vitro</i>	IC <sub>50</sub> * and MIC ranged from 0.044 to 0.049 % and 0.087 to 1.00 % for rocket essential oil, and from 0.049 to 0.282 % and 0.455 to 0.616 % for rosemary essential oil, and from 0.043 to 0.170 % and 0.192 to 0.361 % for tea tree essential oil respectively.	[8]

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Supplementary Table S2. (Continued).

Plant (compound)	Target fungal species	Type of assay	Reduction	References
Clove ( <i>Eugenia caryophyllata</i> ), thyme ( <i>Thymus vulgaris</i> ) and black cumin ( <i>Nigella sativa</i> )	<i>A. flavus</i> <i>F. verticillioides</i>	<i>In vitro</i>	Clove significantly decreased the growth of both tested fungi at all tested concentrations (0.1, 0.5 and 1%); it also resulted in AFs (45.47%) and FBs (33.29%) reduction. Black cumin (1 and 2%) was effective only in suppressing the growth of <i>A. flavus</i> . The main active ingredient in clove oil was eugenol (78.41%).	[9]
<i>Solanum torvum</i> (Torvoside K)	<i>A. flavus</i> <i>F. verticillioides</i>	<i>In vitro/in vivo</i>	MICs ranged from 31.25 to 250 µg/ml <sup>-1</sup> , for <i>A. flavus</i> and <i>F. verticillioides</i> respectively.	[10]
<i>Garcinia kolakola</i> and <i>Azadirachta indica</i>	<i>A. flavus</i> , <i>A. parasiticus</i> ,	<i>In vitro</i>	Inhibition growth was higher by <i>G. kola</i> for both fungi, respectively 77.5% for <i>A. flavus</i> and 54.8% for <i>A. parasiticus</i> , and lowest by <i>A. indica</i> , 35.1% and 30.5% respectively.	[11]
<i>Equisetum arvense</i> and <i>Stevia rebaudiana</i>	<i>A. flavus</i> <i>F. verticillioides</i>	<i>In vitro</i>	Inhibition of growth for both fungi was significant (>99% inhibition at 0.95 a <sub>w</sub> ). AFB <sub>1</sub> and FB <sub>1</sub> presence were not significantly affected.	[12]
<b>Antioxidants/Phenolic compounds</b>				
2-hydroxy-4methoxybenzaldehyde (HMB) from <i>Decalepis hamiltonii</i>	<i>F. verticillioides</i>	<i>In vitro/in vivo</i>	Dose-dependent strong inhibitory activity with MIC value of 100 µg/mL, FB <sub>1</sub> production was completely inhibited at 400 mg/L under <i>in vitro</i> and 750 mg/kg under <i>in vivo</i> .	[13]
Allyl (AITC), phenyl (PITC) and benzyl isothiocyanates (BITC) from cruciferous vegetables	<i>F. verticillioides</i>	<i>In vitro</i>	The mean reduction of FB <sub>2</sub> was 84.9%.	[14]
Allyl isothiocyanate (AITC) from brassica plants	<i>A. parasiticus</i> <i>F. tricinctum</i> <i>F. verticillioides</i> <i>Alternaria alternata</i> <i>F. graminearum</i>	<i>In vitro</i>	AITC treatments at 50, 100 and 500 mL/L inhibited visual growth of all fungal species and kept the production of 12 mycotoxins at undetectable levels (eg.AFB <sub>1</sub> at 72.08 ± 12.70 mg/kg of corn; AFB <sub>2</sub> at 2.14 ± 0.34 mg/kg)	[15]
Ferulic acid	<i>F. verticillioides</i> <i>F. proliferatum</i>	<i>In vitro</i>	The lag phase significantly decreased for both moulds (p ≤ 0.001), However, 10 mM ferulic acid significantly increased (p ≤ 0.001) fumonisin production.	[16]
Tetra-hydro-curcuminoids (THCs) from natural curcuminoids	<i>F. proliferatum</i>	<i>In vitro</i>	Inhibition percentage of fungal growth reached 70% at 13.4 µmol ml <sup>-1</sup> concentration of THCs. FB <sub>1</sub> reduction ranged between 31-37%.	[17]
Budmunchiamine A (BUA) isolated from <i>Albizia amara</i> Pithecolobine (PI) isolated from <i>Albizia saman</i>	<i>A. flavus</i>	<i>In vitro</i>	Inhibitory effect on both <i>A. flavus</i> growth and AFB <sub>1</sub> production by BUA and PI at concentration of 1 mg/mL. MIC ranged from 6.8 to 19.6 mm and 0.015–0.5 mg/mL, respectively for BUA and PI.	[18]
Trans-2-hexenal (T2H)	<i>A. flavus</i>		Significant reduction of fungal populations. Absence of AFB <sub>1</sub> in almost all sets of experiment.	[19]

Minimum inhibitory concentration – MIC; Inhibitory Concentration – IC

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